NOTE

Quantification of Toxic Effects of the Herbicide Metolachlor on Marine Microalgae *Ditylum brightwellii* (Bacillariophyceae), *Prorocentrum minimum* (Dinophyceae), and *Tetraselmis suecica* (Chlorophyceae)

Vinitha Ebenezer and Jang-Seu Ki^{*}

Department of Green Life Science, Sangmyung University, Seoul 110-743, Republic of Korea

(Received March 2, 2012 / Accepted September 17, 2012)

Toxic effects of the herbicide metolachlor (MC) were evaluated for three marine microalgae, *Tetraselmis suecica* (chlorophyte), *Ditylum brightwellii* (diatom), and *Prorocentrum minimum* (dinoflagellate). MC showed a significant reduction in cell counts and chlorophyll *a* levels. Median effective concentration (EC₅₀) was calculated based on chlorophyll *a* levels after a 72-h MC exposure. EC₅₀ values for *T. suecica*, *D. brightwellii*, and *P. minimum* were 21.3, 0.423, and 0.07 mg/L, respectively. These values showed that the dinoflagellate was most sensitive when exposed to the herbicide, at a concentration comparable to freshwater algae, suggesting its potential as an appropriate model organism for ecotoxicity assessments in marine environments.

Keywords: marine microalgae, ecotoxicity assessment, EC₅₀, herbicide, metolachlor

In aquatic environments, microalgae constitute the base of the aquatic food chain. Additionally, they are known for their diversity and sensitivity to environmental changes, thus making them potential candidates for environmental risk assessment studies evaluating toxic contaminants (Stauber and Davies, 2000). Toxicity tests mainly utilize endpoints that measure growth rate, cell density, or chlorophyll content of test species (OECD, 2006). However, the algal sensitivity not only varies among toxicants but also among taxonomic groups (Boyle, 1984). Even the sensitivities to the same toxicant are very different between freshwater algae and marine algae (Sverdrup *et al.*, 2001). Thus, toxicity data employing freshwater algae cannot be used as a surrogate for testing in marine environments and vise versa.

Metolachlor (MC) affects the functioning of membrane

structural components through the impairment of lipid and protein synthesis (Fuerst, 1987), as well as respiration and photosynthesis in plants (Sloan and Camper, 1986). It has been used as a common herbicide for annual weed control in corn, potato fields, and golf courses (US EPA, 2000). This chemical may cause toxicity to aquatic organisms through distributed-source pollution (Lin et al., 1999). The concentration of MC in the aquatic environment was found to range from $5-80 \mu g/L$, and the variation was dependent on season (Cook and Moore, 2008). In addition, the marine environment may be contaminated with various herbicides as a result of spray-drift, run-offs and accidental spills or from freshwater sources (Brock et al., 2006). The toxicity of MC is well documented for freshwater algae (Fairchild et al., 1997; Ma and Liang, 2001; Roubeix et al., 2011) with a selective study in a marine microalgae, Nannochloropsis oculata (Kyriakopoulou et al., 2009). With almost no studies carried out using marine diatoms and dinoflagellates, studies utilizing these marine microalgae as test species are desirable.

In the present study we evaluated the toxic effects of the herbicide MC on three marine microalgae. These included the chlorophyte Tetraselmis suecica, the diatom Ditylum *brightwellii*, and the dinoflagellate *Prorocentrum minimum*, because they have been used previously as test organisms for toxicity studies in marine environments (Canterford and Canterford, 1980; Gerringa et al., 1995; Pérez-Rama et al., 2001; Millán de Kuhn et al., 2006). For our toxicity tests, we obtained microalgal cultures of T. suecica (P-009), D. brightwellii (B-326), and P. minimum (D-127) from the Korea Marine Microalgae Culture Center (Busan, Korea). They were cultured in f/2 medium, and maintained at 20°C and a 12:12 h light:dark cycle with a photon flux density of ca. 65 μ mol photons/m²/s. Exponential growth phase cultures (50 ml) were treated in duplicate with different nominal concentrations (~100 mg/L) of MC (Cat. No. 36163, Sigma, USA), and sub-samples were withdrawn at 0, 12, 24, 48, and 72 h. The concentrations used for toxicity assays were 0.01, 0.05, 0.1, 0.5, 1, 10, 20, and 50 mg/L for D. brightwellii and P. minimum, with additional 75 and 100 mg/L for T. suecica based on previous studies (St-Laurent et al., 1992; Juneau et al., 2001; Kyriakopoulou et al., 2009). Cell counts and chlorophyll a levels were chosen as endpoints to evaluate the toxicity of MC. The 72-h EC₅₀ (median effective

^{*}For correspondence. E-mail: kijs@smu.ac.kr; Tel.: +82-2-2287-5449; Fax: +82-2-2287-0070



Fig. 1. Dose response curves of a chlorophyte, *T. suecica* (A), a diatom, *D. brightwellii* (B) and a dinoflagellate, *P. minimum* (C) based on chlorophyll *a* levels.

concentration) and the percentile inhibition were calculated as recommended in the Organization for the Economic Cooperation and Development (OECD) test guidelines (OECD, 2006). The values of 72-h EC_{50} , including EC_5 , EC_{10} , and EC₂₀, were estimated using a sigmoidal dose-response curve. They were plotted using Origin 8.5 (MicroCal Inc., USA) based on the sigmoidal 4-parameter equation (Teisseyre and Mozrzymas, 2006): Log EC₅₀ = $a + (b - a)/[1 + 10 \times (x - a)/(1 + 10 \times (x - a))/(1 + 10 \times (x - a))/(1 + 10 \times (x - a)))$ $(-c)^{a}$, where a is the response value at zero or minimum asymptote, b is the response value for infinite concentration or maximum asymptote, c is the mid-range point, d is the steepness of the curve or the Hill slope, and x is the dilution coefficient. Dunnett's analysis was carried out to estimate no observable effective concentration (NOEC) and lowest observable effective concentration (LOEC) by using Graphpad InStat (GraphPad Inc., USA). All the data presented are mean values of duplicates; differences between non-treated and treated cultures were tested by one-way analysis of variance (ANOVA) with post hoc Student's Newmann Keul's test in Graphpad InStat (GraphPad Inc., USA).

The toxic effects of MC on the three microalgae were evaluated by measuring cell counts and chlorophyll a levels. Tested microalgae showed marked sensitivities to the herbicide, with patterns depicting dose-dependent reductions in cell counts and chlorophyll a levels. Pearson's correlation coefficient (r) was compared between chlorophyll a and cell counts. T. suecica (0.92, P=0.1247) and P. minimum (0.99, P=0.117) showed a positive correlation between the two endpoints. D. brightwellii, however, showed a negative correlation (-0.827, P=0.0007), because D. brightwellii cells aggregated, thus making it impossible to count individual cells. Hence, we calculated the 72-h EC₅₀ values for the three microalgal test species only using chlorophyll *a* levels (Fig. 1). The 72-h EC₅₀ values for *T. suecica*, *D. brightwellii*, and P. minimum were 21.3±0.2 mg/L, 0.423±0.090 mg/L, and 0.073±0.015 mg/L, respectively (Table 1). As for the threshold effect parameter, we calculated NOEC, LOEC, EC₅, EC₁₀, and EC₂₀ values, which represented the initial concentration of the test chemical that triggers an effect on the test microalgae (Table 1).

Based on literature surveys, we summarized the MC toxicity based on EC_{50} , half maximal inhibitory concentration (IC_{50}), and NOEC values (Table 2). With regard to MC toxicity, most available data were for freshwater algae, excluding the marine eustigmatophyte *Nannochloropsis oculata* (Kyriakopoulou *et al.*, 2009). Our study provided additional toxicity data for MC to marine microalgae represented by three common taxonomic groups, such as chlorophyte (green algae), diatom, and dinoflagellate. Overall, they responded differentially to MC (see Fig. 1), suggesting that the herbicide causes a heterogeneous response to different microalgal groups (Shi *et al.*, 2011).

Upon comparisons of our EC₅₀ data with available literature (Table 2), the chlorophyte T. suecica was most tolerant to MC. In addition, this species was ~50 to ~200 times more tolerant to MC exposure than D. brightwellii and P. minimum, respectively. This was in accordance with earlier studies using metals (Millán de Kuhn et al., 2006; Debelius et al., 2009). D. brightwellii has commonly been used as a test species in toxicity evaluation and bioaccumulation studies (Gerringa et al., 1995). Although this species was observed to be sensitive to metal exposures (Canterford and Canterford, 1980; Gerringa et al., 1995), it displayed moderate toxicity to MC compared to other test species. Interestingly, EC₅₀ values recorded for D. brightwellii (0.423 mg/L) in the present study were comparable to the OECD recommended freshwater diatom, Navicula pelliculosa (0.38 mg/L) (Dobbins et al., 2010). MC was most toxic to the dinoflagellate, P. minimum (72-h EC₅₀=0.073 mg/L). The sub-lethal toxicity response of the dinoflagellate was comparable to the freshwater chlorophytes, Ankitrodesmus falcatus, Chlorella pyrenoidosa, and Pseudokirchneriella subcapitata (formerly known as *Selenastrum capricornutum*); however, it was more sensitive than the chlorophytes, Scenedesmus spp. and Pediastrum biwae (Table 2).

Additionally, the toxicity of MC to marine microalgae was compared to other herbicides and insecticides. For example, the EC_{50} values of the herbicide triazine to marine diatoms

Table 1. The effective concentration after 72 h exposure of metolachlor to three microalgae									
Species	NOEC (mg/L)	LOEC (mg/L)	$EC_5(mg/L)$	EC10 (mg/L)	EC ₂₀ (mg/L)	EC_{50} (mg/L)	95% confidence limits		
T. suecica	$0.088 {\pm} 0.001$	$0.154 {\pm} 0.030$	$0.690 {\pm} 0.008$	3.427 ± 0.660	$10.39 {\pm} 0.001$	21.3±0.020	19.85-22.19		
D. brightwellii	0.0025	0.001	0	0.008 ± 0.0002	$0.085 {\pm} 0.001$	0.423 ± 0.090	0.254-0.625		
P. minimum	0	0	$0.001 {\pm} 0.0004$	0.012 ± 0.001	0.035 ± 0.005	0.073±0.015	0.068-0.084		

138 Ebenezer and Ki

	te metolachior to common freshwater a			
Taxonomic position	Species	$EC_{50}/IC_{50}/NOEC (mg/L)$	References	
Freshwater species				
Chlorophyceae	Ankitrodesmus falcatus	0.096	Juneau <i>et al.</i> (2001)	
Chlorophyceae	Chlorella pyrenoidosa	0.068-0.152	Liu and Xiong (2009)	
Chlorophyceae	Psedokirchneriella subcapitata	0.037-0.084	Fairchild et al. (1997), Juneau et al. (2001)	
Chlorophyceae	Pediastrum biwae	0.330	Juneau <i>et al</i> . (2001)	
Chlorophyceae	Scenedesmus acutus	0.1	St-Laurent <i>et al.</i> (1992)	
	Scenedesmus vacuolatus	$0.156 - 0.598^{a}$	Kotrikla <i>et al.</i> (1999)	
Bacillariophyceae	Navicula pelliclosa	0.38	Dobbins et al. (2010)	
Nostocaceae	Anabaena cylindrical	4.0^{b}	St-Laurent <i>et al.</i> (1992)	
Chroococcales	Microcystis aeruginosa	0.073	Juneau <i>et al</i> . (2001)	
Nannoplankton	-	0.117	Juneau <i>et al</i> . (2001)	
River periphytic diatoms	-	0.005-0.03	Roubeix <i>et al.</i> (2011)	
Marine Species				
Eustigmatophyceae	Nannochloropsis oculata	10.5	Kyriakopoulou et al. (2009)	
^a IC ₅₀ value ^b NOEC				

Cyclotella gamma and Chaetoceros sp. were 0.494 mg/L and 0.043 mg/L, respectively (Tang et al., 1997; Debelius et al., 2008). Endosulfan (insecticide) was very toxic to the dinoflagellate, *P. minimum*, for which the EC₅₀ value was 0.025 mg/L (unpublished data). EC₅₀ of the insecticide, cypermethrin to the marine dinoflagellate, Scrippsiella trochoidea was reported as 0.205 mg/L (Wang et al., 2012). Moreover, the marine algae, D. brightwellii and P. minimum were more sensitive to MC than freshwater and marine invertebrates; for example, 24-h LC₅₀ values of MC to the freshwater crustacean, Daphnia magna and marine crustacean, Artemia franciscana were 9.5 mg/L and 168 mg/L, respectively (Kyriakopoulou et al., 2009). These results suggest that marine diatoms and dinoflagellates may be very sensitive to herbicide exposures (Védrine et al., 2003), and can be used as model test organisms in aquatic toxicity assessment studies.

This work was supported by the "Eco-innovation project" of the Ministry of Environment, Korea.

References

- Boyle, T.P. 1984. Effect of environmental contaminants on aquatic algae, pp. 237–256. *In* Shubert, L.E. (ed.), Algae as Ecological Indicators. Academic Press, New York, N.Y., USA.
- Brock, T.C.M., Arts, G.H.P., Maltby, L., and van den Brink, P.J. 2006. Aquatic risks of pesticides, ecological protection goals, and common aims in European Union legislation. *Intgr. Environ. Manag. Assess.* 2, 20–46.
- **Canterford, G.S. and Canterford, D.R.** 1980. Toxicity of heavy metals to the marine diatom *Ditylum brightwellii* (West) Grunow: correlation between toxicity and metal speciation. *J. Mar. Biol. Ass. U.K.* **60**, 227–242.
- Cook, M.E. and Moore, P.A. 2008. The effects of the herbicide metolachlor on agonistic behavior in the Crayfish, *Orconectes rusticus*. Arch. Environ. Contam. Toxicol. 55, 94–102.
- Debelius, B., Forja, J.M., Del Valls, A., and Lubian, L.M. 2008. Effect of linear alkyl benzene sulfonate (LAS) and atrazine on marine microalgae. *Mar. Poll. Bull.* 57, 559–568.

Debelius, B., Forja, J.M., Del valls, A., and Lubian, L.M. 2009.

Toxicity and bioaccumulation of copper and lead in five marine microalgae. *Ecotoxicol. Environ. Saf.* **72**, 1503–1513.

- **Dobbins, L., Lewis, M., Sankula, S., and Thursby, G.** 2010. Exploration of methods for characterising effects of chemical stressors to aquatic plants. http://water.epa.gov. (Last accessed 20 Jan. 2012).
- Fairchild, J.F., Ruessler, D.S., Haverland, P.S., and Carlson, A.R. 1997. Comparative sensitivity of Selenastrum capricornutum and Lemna minor to sixteen herbicides. Arch. Environ. Contam. Toxicol. 32, 353–357.
- Fuerst, E.P. 1987. Understanding the mode of action of chloroacetamide and thiocarbamate herbicides. *Weed Technol.* 1, 270– 277.
- Gerringa, L.J.A., Rijstenbil, J.W., Poortvleit, T.C.W., van Drie, J., and Schot, M.C. 1995. Speciation of copper and responses of the marine diatom *Ditylum brightwellii* upon increasing copper concentrations. *Aquat. Toxicol.* 31, 77–90.
- Juneau, P., Dewez, D., Matsui, S., Kim, S.-G., and Popovic, R. 2001. Evaluation of different algal species sensitivity to mercury and metolachlor by PAM-fluorometry. *Chemosphere* 45, 589–598.
- Kotrikla, A., Gatidou, G., and Lekkas, T.D. 1999. Toxic effects of atrazine, deethyl-atrazine, deisopropyl-atrazine and metolachlor on *Chlorella fusca var-fusca. Global Nest: the Int. J.* 1, 39–45.
- Kyriakopoulou, K., Anastasiadou, P., and Machera, K. 2009. Comparative toxicities of fungicide and herbicide formulations on freshwater and marine species. *Bull. Environ. Contam. Toxicol.* 82, 290–295.
- Lin, Y.-J., Karuppiah, M., Shaw, A., and Gupta, G. 1999. Effect of simulated sunlight on atrazine and metolachlor toxicity of surface waters. *Ecotoxicol. Environ. Saf.* 43, 35–37.
- Liu, H. and Xiong, M. 2009. Comparitive toxicity of racemic metolachlor and S-metolachlor to *Chlorella pyrenoidosa*. Aquat. Toxicol. 93, 100–106.
- Ma, J. and Liang, W. 2001. Acute toxicity of 12 herbicides to the green algae Chlorella pyrenoidosa and Scenedesmus obliqus. Bull. Environ. Contam. Toxicol. 67, 347–351.
- Millán de Kuhn, M., Streb, C., Breiter, R., Richter, P., Neeße, T., and Häder, D.P. 2006. Screening for unicellular algae as possible bioassay organisms for monitoring marine water samples. *Water Res.* 40, 2695–2703.
- **OECD.** 2006. OECD Guidelines for the testing of chemicals. Freshwater algal and cyanobacteria growth inhibition test. 201. OECD Publications, Paris, France.

- Pérez-Rama, M., Herrero López, C., Abalde Alonso, J., and Torres Vaamonde, E. 2001. Class III metallothioneins in response to cadmium toxicity in the marine microalga *Tetraselmis suecica* (Kylin) Butch. *Environ. Toxicol. Chem.* 20, 2061–2066.
- Roubeix, V., Mazzella, N., Mechin, B., Coste, M., and Delmas, F. 2011. Impact of the herbicide metolachlor on river periphytic diatoms: experimental comparison of descriptors at different biological organization levels. *Ann. Limnol. Int. J. Lim.* **47**, 1–11.
- Shi, X.L., Lepère, C., Scanlan, D.J., and Vaulot, D. 2011. Plastid 16S rRNA gene diversity among eukaryotic picophytoplankton sorted by flow cytometry from the South Pacific Ocean. *PLoS ONE* 6, e18979.
- Sloan, M.E. and Camper, N.D. 1986. Effects of alachlor and metolachlor on cucumber seedlings. *Environ. Exp. Bot.* 26, 1–7.
- Stauber, J.L. and Davies, C.M. 2000. Use and limitations of microbial bioassays for assessing copper availability in the aquatic environment. *Environ. Rev.* 8, 255–301.
- St-Laurent, D., Blaise, C., Macquarrie, P., Scroggins, R., and Trottier,
 B. 1992. Comparative assessment of herbicide phytotoxicity to Selenastrum capricornutum using microplate and flask bioassay

procedures. Environ. Toxicol. 7, 35-48.

- Sverdrup, L.E., Källqvist, T., Kelley, A.E., Fürst, C.S., and Hagen, S.B. 2001. Comparative toxicity of acrylic acid to marine and freshwater microalgae and significance for environmental effects assessments. *Chemosphere* 45, 653–658.
- Tang, F.-X., Hoagland, K.D., and Siegfried, B.D. 1997. Differential toxicity of atrazine to selected freshwater algae. *Bull. Environ. Contam. Toxicol.* 59, 631–637.
- Teisseyre, A. and Mozrzymas, J.W. 2006. The inhibitory effect of copper ions on lymphocyte KVI.3 potassium channels. *J. Physiol. Pharmacol.* **57**, 301–314.
- US EPA. 2000. Equivalency of pesticides metolachlor and S-metolachlor with respect to ground water contamination. Federal Register Document, Volume 65, No. 36. http://www.gpo.gov/
- Védrine, C., Leclerc, J.C., Durrieu, C., and Tran-minh, C. 2003. Optical whole-cell biosensor using *Chlorella vulgaris* designed for monitoring herbicides. *Biosens. Bioelectron.* 18, 457–463.
- Wang, Z.-H., Nie, X.-P., Yue, W.-J., and Li, X. 2012. Physiological responses of three marine microalgae exposed to cypermethrin. *Environ. Toxicol.* 27, 563–572.